

trum⁵ included singlets at δ 0.65 (3 H) for the C-19 methyl, 0.92 (3 H) for the C-18 methyl, 1.58 (3 H) for $\text{CH}_3\text{C}=\text{C}$, and 2.13 (3 H) for the C-21 methyl. Crystallization from pentane gave material mp 88–89°, which contained <5% of the 17α form by vpc.

The tetracyclic ketone **12** was submitted to ozonolysis to give the triketone **13** which, without purification, was treated so as to induce intramolecular aldol condensation. Preparative tlc of the product on silica gel (40% EtOAc in pentane) gave (45% overall yield from **12**) a crystalline product which contained 85% of the 17β (**14**) and 15% of the 17α isomer. Two recrystallizations gave *dl*-progesterone, mp 182–185°, undepressed on admixture with authentic material,¹⁰ mp 182–185.5°. The solution ir, nmr, and mass spectra of the former specimen were identical with the respective spectra of the latter as well as of natural progesterone. It is to be noted that many of the yields recorded in this paper have not yet been optimized.

Acknowledgments. We are indebted to the National Institutes of Health and the National Science Foundation for support of this research.

(10) W. S. Johnson, J. A. Marshall, J. F. W. Keana, R. W. Franck, D. G. Martin, and V. J. Bauer, *Tetrahedron, Suppl.*, **8** (2), 541 (1966).

William S. Johnson,* Michael B. Gravestock, Brian E. McCarry
Department of Chemistry, Stanford University
Stanford, California 94305
Received May 21, 1971

A Correlation of Some Structural Parameters of Pyrimidine Nucleosides. A Nuclear Magnetic Resonance Study

Sir:

A number of workers have drawn attention to the apparent correlation between the nature of the sugar-ring puckering and the sugar-base torsion angle (ϕ_{CN}) of crystalline nucleosides as determined by X-ray studies.^{1–3} Prestegard and Chan⁴ described correlations between the chemical shift of the H₆ hydrogen and the $J_{1'2'}$ coupling constant of pyrimidine nucleosides which suggest that this torsion angle–ring puckering relationship may exist in aqueous solution as well. Wilson and Rahman⁵ pointed out that there may in some cases be a mutual relation between ring puckering and the nucleoside conformation about the exocyclic C₄–C_{5'} bond. It was to investigate the possibility of the existence of this latter relationship in aqueous solution that the present pmr study of a variety of pyrimidine nucleosides was initiated. Our data demonstrate that the gauche–gauche conformation is favored if the ribose, or deoxyribose, is in the C_{3'} endo conformation and less favored if the sugar is puckered C_{2'} endo.

The nucleosides were purchased from Calbiochem and the Sigma Chemical Co.; 0.1 M aqueous (D₂O) solutions containing 0.05 M internal reference (2,2,3,3-tetradeuterio-3-(trimethylsilyl)propionic acid sodium

salt) were lyophilized three times. Spectra were obtained at 220 MHz (23°) and analyzed using LAME.⁶ The spectral assignments were carried out by comparison of the shift data in the ribose and deoxyribose series with consideration of published work.^{4,7} Some of the data for the correlation were taken from the literature. The literature sources are indicated in the caption of Figure 2.

The $J_{1'2'}$ and $J_{3'4'}$ couplings provide a means of monitoring the average conformation of the sugar ring. They can be related, *via* the Karplus equation,⁸ to the dihedral angles $\phi_{1'2'}$ and $\phi_{3'4'}$ in the relevant HCC'H' fragments (Figure 1). Smith and Jardetzky⁹ have estimated the dihedral angles and corresponding coupling constants for a number of possible puckered conformations of ribose and deoxyribose rings. From their table we can make the following qualitative predictions: (a) with increasing proportion of the C_{3'} endo (and/or C_{2'} exo) conformation $\phi_{3'4'}$ and $J_{3'4'}$ increase whereas $\phi_{1'2'}$ and $J_{1'2'}$ decrease and (b) the reverse of (a) is the case for increasing proportion of the C_{2'} endo (and/or C_{3'} exo) conformation.¹⁰

Newman projection formulas along the C₄–C_{5'} bond are shown in Figure 1. Clearly, in the gauche–gauche rotamer in which the 5' hydroxyl lies above the sugar ring the magnitudes of the vicinal coupling constants $J_{4'5'B}$ and $J_{4'5'C}$ ¹¹ are predicted by the Karplus equation to be small (*ca.* 2 Hz), since the coupled nuclei are in a gauche configuration ($\phi_{4'5'B}$ and $\phi_{4'5'C} = 60^\circ$). However, if the 5' hydroxyl lies off the ring in either the trans–gauche or gauche–trans rotamer, the 4' hydrogen is trans to one methylene hydrogen ($\phi = 180^\circ$) and the corresponding vicinal J is predicted to be large (*ca.* 10 Hz). Thus any perturbation resulting in a decrease in the proportion of the gauche–gauche rotamer should be manifest in an increase in the observed sum of $J_{4'5'B} + J_{4'5'C}$.¹²

In Figures 2 and 3 we have plotted this sum *vs.* $J_{3'4'}$ and $J_{1'2'}$, respectively. Clearly, a good correlation exists for the nucleosides indicated by X. Further, we note that the sum decreases smoothly with increasing $J_{3'4'}$, but increases with increasing $J_{1'2'}$. Both trends are consistent with a correlated decrease in the gauche–gauche population with an increasing proportion of the C_{2'} endo (and/or C_{3'} exo) conformation in this series of ribo- and deoxyribonucleosides. The molecules on the correlation lines have the following common structural features: (a) they are β anomers, (b) they have a C–H and C=O at the ortho positions of the base, and (c) they most certainly exist in their anti conformation in solution.^{13–26}

(6) C. W. Haigh and J. M. Williams, *J. Mol. Spectrosc.*, **32**, 398 (1969).

(7) R. U. Lemieux, *Can. J. Chem.*, **39**, 116 (1961).

(8) M. Karplus, *J. Chem. Phys.*, **30**, 11 (1959).

(9) M. Smith and C. D. Jardetzky, *J. Mol. Spectrosc.*, **28**, 70 (1968).

(10) Endo means the atom is located on the same side of the plane defined by C_{1'}, O_{1'}, and C_{4'} as the C₄–C_{5'} bond. Exo means that it is found on the opposite side.

(11) In the spectral analyses the 4' and 5' hydrogens are treated as an ABC subsystem, hence the subscripts B and C.

(12) We realize that the actual rotamers about the C₄–C_{5'} bond may be somewhat distorted from the $\phi = 60^\circ$ values due to O–O repulsion. Qualitatively identical trends are predicted on a distorted rotamer model. Further, since an absolute assignment of the magnetically nonequivalent methylene hydrogens cannot be made at this time, the sum of the two vicinal couplings must be used as a qualitative measure of the relative rotamer populations.

(13) B. J. Blackburn, A. A. Grey, I. C. P. Smith, and F. E. Hruska, *Can. J. Chem.*, **48**, 2866 (1970).

(1) A. E. V. Haschmeyer and A. Rich, *J. Mol. Biol.*, **27**, 369 (1967).

(2) M. Sundaralingam, *Biopolymers*, **7**, 821 (1969).

(3) A. V. Lakshminarayanan and V. Sasisekharan, *Biochim. Biophys. Acta*, **204**, 49 (1970).

(4) J. H. Prestegard and S. I. Chan, *J. Amer. Chem. Soc.*, **91**, 2843 (1969).

(5) H. R. Wilson and A. Rahman, *J. Mol. Biol.*, **56**, 129 (1971).

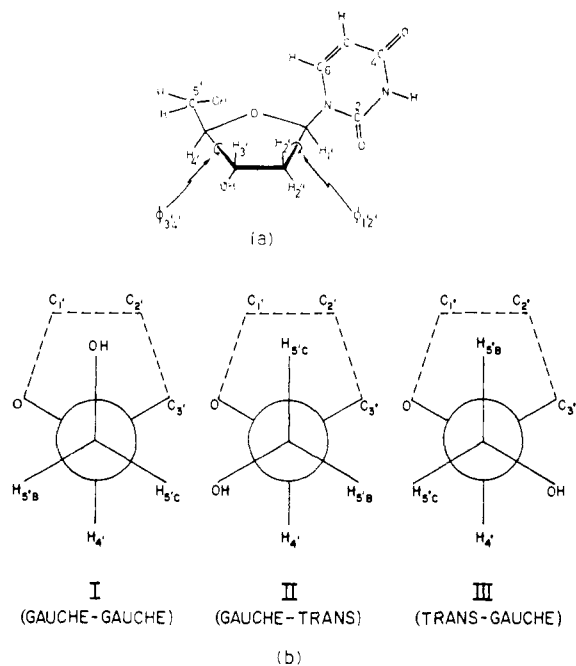


Figure 1. (a) Structure of 2'-deoxyuridine indicating the dihedral angles $\phi_{1/2'}$ and $\phi_{3/4'}$; (b) possible rotational isomers around the $C_4'-C_5'$ bond of a nucleoside.

It is interesting to note that the approximate data for poly-U¹⁹ fall near to both correlation plots. However, points for the α anomer of pseudouridine do not correlate. Further, the points for O and β -RCA (in which a bulky keto group lies in close proximity to the ribose ring^{15,16,26}) and AU and AC (in which an aza nitrogen replaces the C_6-H group) lie distant from the curves. It is apparent that in the compounds which fall off the line the C_2' endo pucker is not probable. A possible rationalization of the deviations may be based on anomalous nonbonded interactions between such keto groups or aza nitrogens and the sugar ring.

X-Ray data¹⁻³ indicate that the gauche-gauche conformation is most commonly observed for crystalline nucleosides with either the C_2' endo or C_3' endo structures. However, in the two instances with C_3' exo sugars (thymidine²⁷ and deoxyadenosine²⁸) the gauche-

- (14) F. E. Hruska, K. K. Ogilvie, A. A. Smith, and H. Wayborn, *Can. J. Chem.*, **49**, 2449 (1971).
- (15) F. E. Hruska, *J. Amer. Chem. Soc.*, **93**, 1795 (1971).
- (16) H. Dugas, B. J. Blackburn, R. K. Robins, R. Deslauriers, and I. C. P. Smith, *ibid.*, **93**, 3468 (1971).
- (17) F. E. Hruska, A. A. Grey, and I. C. P. Smith, *ibid.*, **92**, 214, 4088 (1970).
- (18) A. A. Grey, I. C. P. Smith, and F. E. Hruska, *ibid.*, **93**, 1765 (1971).
- (19) S. I. Chan and G. P. Krieshman, *ibid.*, **92**, 1102 (1970).
- (20) T. R. Emerson, R. J. Swan, and T. L. V. Ulbricht, *Biochemistry*, **6**, 834 (1967).
- (21) M. P. Schweizer, A. D. Broom, P. O. P. Ts'ao, and D. P. Hollis, *J. Amer. Chem. Soc.*, **90**, 1042 (1968).
- (22) D. W. Miles, W. H. Inskip, M. J. Robins, M. W. Winkley, R. K. Robins, and H. Eyring, *ibid.*, **92**, 3872 (1970).
- (23) A. Rahman and H. R. Wilson, *Nature (London)*, **225**, 64 (1970).
- (24) C. L. Coulter and S. W. Hawkinson, *Proc. Nat. Acad. Sci. U. S.*, **63**, 1359 (1969).
- (25) M. P. Schweizer, J. T. Witkowski, and R. K. Robins, *J. Amer. Chem. Soc.*, **93**, 277 (1971).
- (26) F. E. Hruska, *Can. J. Chem.*, **49**, 2111 (1971).
- (27) H. R. Wilson, A. Rahman, and P. Tollin, *J. Mol. Biol.*, **46**, 585 (1969).
- (28) D. G. Watson, D. J. Sutor, and P. Tollin, *Acta Crystallogr.*, **19**, 111 (1965).

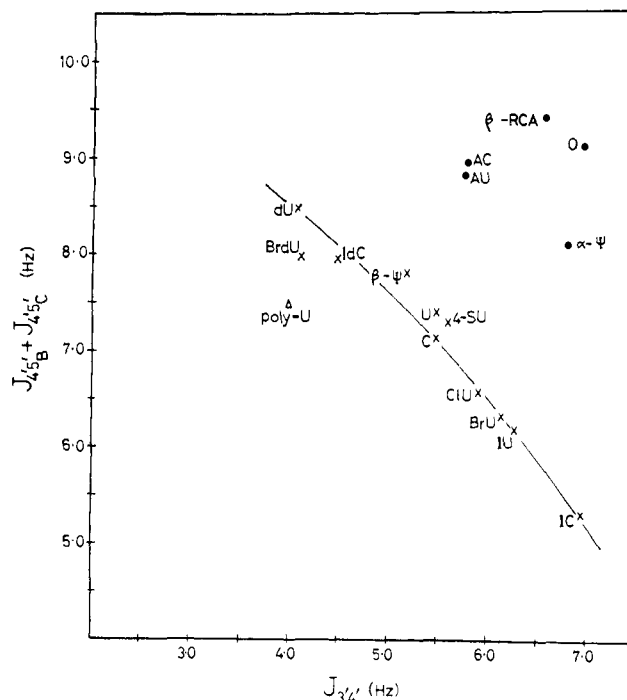


Figure 2. Plot of $J_{4/5'B} + J_{4/5'C}$ vs. $J_{3/4'}$ for a series of pyrimidine nucleosides. Abbreviations: U, CIU, BrU, and IU for uridine^{1,3} and its 5-substituted halo derivatives; dU and BrdU for the analogous deoxy derivatives; C, IC, and IdC for cytidine, 5-iodo-, and 5-iododeoxycytidine; 4-SU, 4-thiouridine;¹⁴ O, orotidine;¹⁵ β -RCA, 1-(B-D-ribofuranosyl)cyanuric acid;¹⁶ AU, 6-azauridine; AC, 6-azacytidine; α - ψ and β - ψ , α - and β -pseudouridine;^{17,18} poly-U, polyuridylic acid.¹⁹ References where given are to the data source.

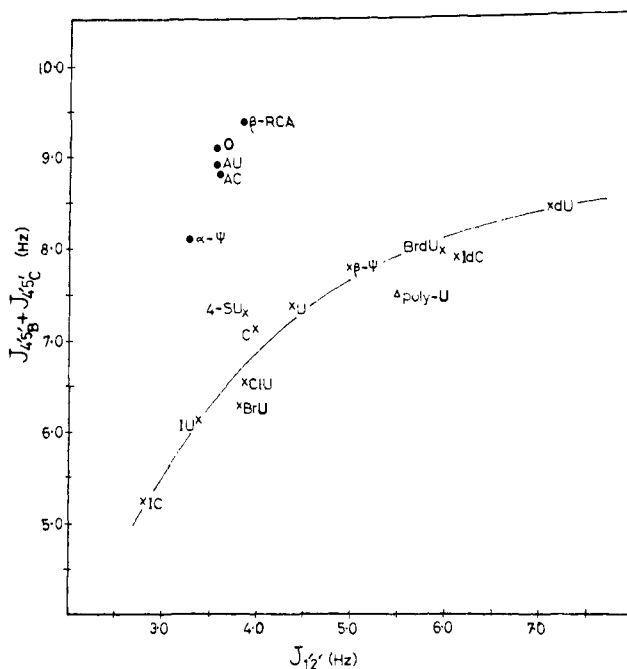


Figure 3. Plot of $J_{4/5'B} + J_{4/5'C}$ vs. $J_{1/2'}$ for a series of pyrimidine ribo- and deoxyribonucleosides. Abbreviations as in Figure 2.

trans conformation is observed. In addition, Rahman and Wilson²³ have noted that in crystalline 5-iodouridine the molecules exist in two nonequivalent environments, those in one exhibiting a C_3' endo-gauche-gauche con-

formation, those in the other, a C_{2'} endo-trans-gauche conformation.²⁹

Acknowledgment. We are grateful to Mr. R. Wasylihen for his help with the computer analyses and Dr. J.B. Rowbotham for his adaptation of a plotting routine for LAME and to the Faculty of Graduate Studies of the University of Manitoba for financial support. J. G. D. is grateful for the award of a Manitoba Fellowship. This work was supported by a grant from the National Research Council of Canada. We thank Dr. A. A. Grey of the Ontario Research Foundation for obtaining the 220-MHz spectra. We thank a referee for suggesting the inclusion of the statement regarding the low probability of the C_{2'} endo conformation for those nucleosides which do not correlate.

(29) NOTE ADDED IN PROOF. A correction (*ca.* 1 Hz) for the effect of the electronegativity differential between 2'-H and 2'-OH should be applied to the $J_{1'2'}$ values of dU, IdC, and BrdU. A much smaller correction (<0.5 Hz) might be required for the corresponding $J_{3'4'}$ values. These corrections would shift the three points to the left in Figure 3 and to the right in Figure 2 but do not affect our qualitative conclusions. Further, since submission of this article we have analyzed spectra for thymidine (T), dihydrouridine (diHU), and arabinouridine (aU). Data for T and diHU correlate, although the latter does not have feature b in common with the correlating nucleosides. However, aU does not correlate; this suggests that the stereoconfiguration of the sugar is important insofar as the correlation is concerned.

F. E. Hruska,* A. A. Smith, J. G. Dalton
Chemistry Department, The University of Manitoba
Fort Garry, Manitoba, Canada
Received May 24, 1971

Emission from Tungsten Carbonyl Complexes

Sir:

Dissociative decay is a major deactivation pathway for electronic excited states of metal carbonyls.¹ Other facile nonradiative processes include reactions of coordinated ligands other than CO¹⁻³ and internal conversion where photoreaction quantum yields are less than unity.⁴⁻⁶ In this communication we report the observation of emission from W(CO)₅(L) compounds

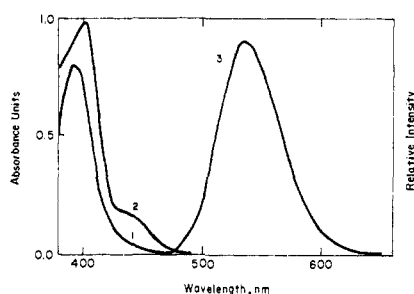


Figure 1. Room temperature absorption spectra of Mo(CO)₅(cyclohexylamine), curve 1, and W(CO)₅(cyclohexylamine), curve 2. Spectra recorded for $\sim 10^{-4}$ M isoctane solutions using 1-cm path cells. Curve 3 shows 77°K emission from W(CO)₅(cyclohexylamine). No emission could be seen from Mo(CO)₅(cyclohexylamine).

(1) E. Koerner von Gustorf and F. W. Grevels, *Fortschr. Chem. Forsch.*, **13**, 366 (1969).

(2) M. Wrighton, G. S. Hammond, and H. B. Gray, *J. Amer. Chem. Soc.*, **93**, 3285 (1971).

(3) W. Jennings and B. Hill, *ibid.*, **92**, 3199 (1970).

(4) A. P. Garratt and H. W. Thompson, *J. Chem. Soc.*, 1817 (1934).

(5) W. Strohmeier and D. von Hobe, *Chem. Ber.*, **94**, 2031 (1961).

(6) W. Strohmeier and D. von Hobe, *Z. Phys. Chem. (Frankfurt am Main)*, **34**, 393 (1962).

where L is a ketone, ether, amine, or pyridine. This is the first report of a radiative deactivation pathway for metal carbonyl complexes.

Preparation of the W(CO)₅(L) compounds was achieved by photolysis of W(CO)₆ in the presence of the entering ligand.¹ Infrared spectra in the carbonyl region are in agreement with previously published data.⁷ The uv-vis spectra of the complexes studied are similar, with a prominent feature being an intense absorption in the vicinity of 400 nm. Room-temperature emission from the complexes could not be detected, but upon cooling to 77°K all of the complexes exhibited an emission maximum between 510 and 545 nm.⁸ The emission spectra show little or no structure and there is some overlap of the emission and absorption bands. Figure 1 shows typical emission and absorption spectra. Table I gives emission maxima as well as low-energy

Table I. Emission and Absorption Maxima for Some M(CO)₅(L) Complexes

Compound	Absorption max, nm	Emission max, nm
W(CO) ₅ (NEt ₃)	465, 428, 402	533
Mo(CO) ₅ (NEt ₃)	393	c
W(CO) ₅ (NHEt ₂)	438, 402	533
Mo(CO) ₅ (NHEt ₂)	393	c
W(CO) ₅ (cyclohexylamine)	438, 402	533
Mo(CO) ₅ (cyclohexylamine)	393	c
W(CO) ₅	436 ^a	533
W(CO) ₆	314, 334, 353 ^b	c
Mo(CO) ₆	313, 332, 347 ^b	c
W(CO) ₅ (acetone)	406, 450	538
W(CO) ₅ (ethyl ether)	418, 456	533
Mo(CO) ₅ (ethyl ether)	407	c
W(CO) ₅ (pyridine)	440, 385	510
W(CO) ₅ (S-Et ₂)	412, 386	545
Mo(CO) ₅ (S-Et ₂)	377	c

^a Reference 9. ^b Reference 13. ^c None detectable.

absorption maxima for a number of W(CO)₅(L) species

We have also found weak emission from W(CO)₅ itself. Tungsten pentacarbonyl was produced in a methylcyclohexane glass at 77°K by photolysis of W(CO)₆ as previously reported.^{9,10} The 533-nm emission, however, could not be detected until the excess CO was removed. The lack of emission before removing the trapped CO is probably related to the fact that regeneration of W(CO)₅ can be achieved *via* long-wavelength photolysis of the [W(CO)₅ + CO] in the low-temperature glass.¹¹ The CO in the cage may be weakly interacting as an O-bonded ligand or in a π-type configuration. We have not been able to detect emission from complexes with L = olefins or acetonitrile.

(7) "Organic Synthesis *via* Metal Carbonyls," Vol. I, I. Wender and P. Pino, Ed., Wiley, 1968, and references cited therein.

(8) Emission spectra were obtained by using an Aminco-Bowman spectrophotofluorometer or the optical detection system of a Cary 17. Excitation spectra were obtained with the Aminco instrument, and excitation maxima agreed well with absorption maxima for the compounds. Relative emission intensities were estimates, the emission spectra being obtained by exciting at 400 nm. All emission spectra were obtained at 77°K.

(9) M. A. Graham, A. J. Rest, and J. J. Turner, *J. Organometal. Chem.*, **24**, C54 (1970).

(10) (a) I. W. Stolz, G. R. Dobson, and R. K. Sheline, *J. Amer. Chem. Soc.*, **84**, 3589 (1962); (b) I. W. Stolz, G. R. Dobson, and R. K. Sheline, *ibid.*, **85**, 1014 (1963).

(11) M. Wrighton, G. S. Hammond, and H. B. Gray, unpublished observations.